## **Vascular Disruption Effects of MPC-6827**

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## ABSTRACT

MPC-6827 is an investigational new drug that has recently completed Phase I clinical trials in advanced primary and metastatic tumors. MPC-6827 has been previously reported to potently inhibit tumor cell growth and survival in vitro and in vivo, with activity in xenograft models of mouse melanoma and human cancers of the ovary, breast, prostate, colon and pancreas. MPC-6827 has also been shown to compete with colchicine for binding to β-tubulin and to inhibit the formation of microtubules in vitro and in cell culture. We have recently begun to evaluate the potential of MPC-6827 to act as a vascular disrupting agent (VDA). VDAs target the tumor neovascular endothelium leading to disruption of tumor blood supply and subsequ tumor cell necrosis. In cell culture, MPC-6827 induced cytotoxicity in several primary endothelial cell types, including human umbilical vein, arterial, and microvessel endothelial cells, with IC50 values from 1.8 to 3.2 nM. In human OVCAR-3 ovarian carcinoma xenografts in nude mice, a single intravenous treat of MPC-6827 at 10 mg/kg induced dramatic vascular disruption and tumor cell necrosis within 24 hours. Based on these preliminary results, we conclude that the anti-tumor effects are likely to be mediated, at least in part, through a mechanism involving vascular disruption. We are currently studying the effects in the OVCAR-3 xenograft model in more detail and investigating other xenograft models.

Data obtained since submission of abstract: VDAs have been limited in their efficacy due to the observation that the rim of the tumor survives treatment and will subsequently regrow. Therefore, a combination of standard cytotoxic therapy with a VDA has been proposed to increase efficacy. We have recently tested a single, suboptimal dose of MPC-6827 in combination with a single dose of carboplatin and found synergistic inhibition of OVCAR-3 human ovarian carcinoma xenograft growth. With the addition of these new findings to this poster, the authors have elected not to show the *in vitro* endothelial cell cytoxicity data cited in the original abstract.

## **METHODS**

For OVCAR-3 xenografts, female Crl:Nu/Nu-nuBR mice (Charles River Labs, Wilmington, MA) were implanted subcutaneously with 107 cells in the right flank Animals were housed by groups in Positive Individual Ventilation cages in flatbottom cages with no more than ten mice per cage. Environmental controls were set to maintain a temperature between 65 and 75 °F with a relative humidity of 30-70% in a 12:12 hour light:dark cycle. Animals were fed and watered ad libitum. Tumors were allowed to grow to approximately 100 mm3 (Study 1) or 380 mm3 (Study 3) and then mice were placed into test groups (N = 10). The mice were observed daily for mortality and signs of toxicity. Tumor growth was monitored externally using a caliper and volumes calculated using the formula  $[p/6 (width^2 x length)]$ , in which width represents the smaller tumor diameter. Volume values were converted to percentage of original tumor size and group values averaged. Studies were complete when the first animal achieved a tumor volume > 1500 mm<sup>3</sup>. Statistical analysis of variance with unadjusted pair wise comparison was performed using SAS software (Cary, NC). These studies conformed to the recommendations set forth in the U.S. Public Health Service Policy on Humane Care and Use of Laboratory Animals,1

In Study 1, animals were treated with vehicle, 7.5 mg/kg MPC-6827 qwk x 3 *i.v.* or 30 mg/kg carboplatin qd x 5 *i.p.* In Study 2, mice with OVCAR-3 kenografts were treated with vehicle, combretisatatin A-4 phosphate or MPC-6827. Animals were sacrificed 24 h later and tumors were fixed in formalin. Tissue sections were embedded in paraffin for storage, prior to Hematoxylin and Eosin Y (H & E) stainin using standard techniques 2 in Study 3, OVCAR-3 kenograft hearing mice were treated with a suboptimal single dose of 5 mg/kg MPC-6827 *i.v.* followed 4 h later by 60 mg/kg carboplatin *i.p.*, or 60 mg/kg carboplatin *i.p.* followed 4 h later by 5 mg/kg MPC-6827 *i.v.* 

<sup>1</sup> The U.S. Public Health Service Policy on Humane Care and Use of Laboratory Animals, Office of Laboratory Animal Welfare, National Institutes of Health, Department of Health and Human Services Bethesda, MD 20892-7982
<sup>2</sup> Luna, L. Manual of Histoloeic Staining Methods of the Armed Forces Institute of Patholoev. 3rd

Edition. New York: McGraw-Hill Publisher; 1968.

